

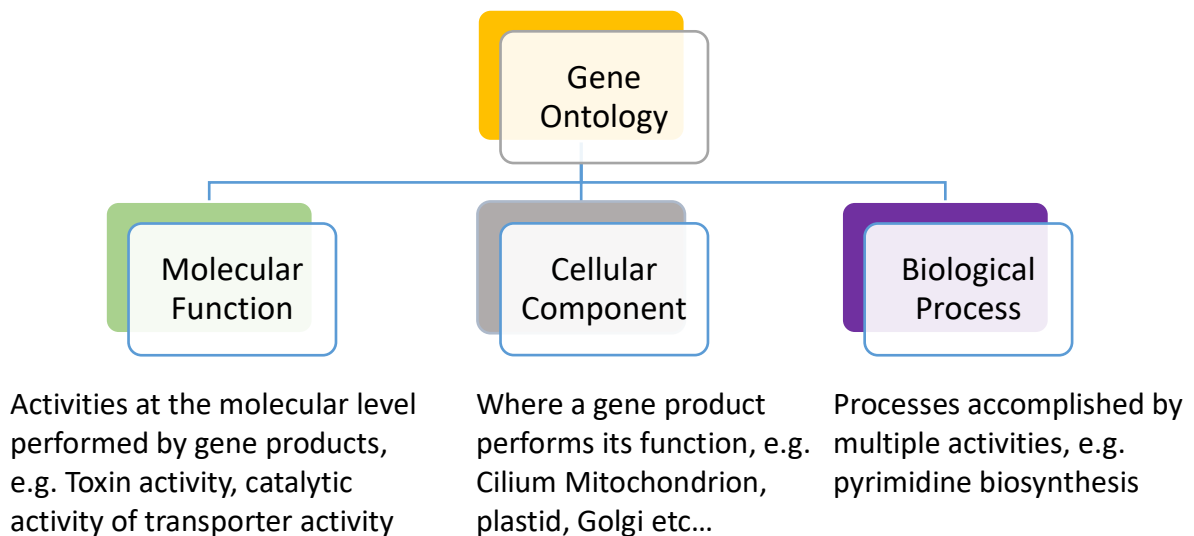
# Gene Ontology (GO) Enrichment

## Learning objectives:

- Run a GO enrichment analysis
- Explore GO enrichment results

## Background:

The gene ontology describes the knowledge of biological sciences and divides this knowledge into three broad categories: Molecular function, cellular component and biological process.



To learn more about Gene Ontology, please visit:  
<http://geneontology.org/docs/ontology-documentation/>

A gene can be assigned a GO term either manually (by an annotator evaluating experimental evidence) or computationally (based on the GO terms of genes that share sequence or functional domains). These GO terms can be used to test whether your set of genes are enriched for a molecular function, cellular component, or biological process.

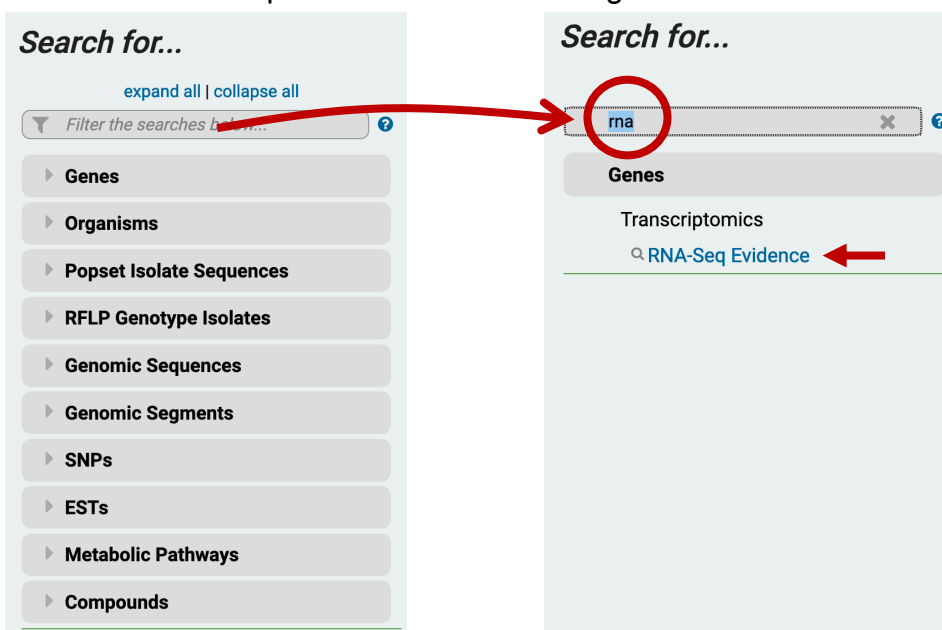
**For example:** Does my list of genes have an over-representation of specific GO terms compared to the rest of the genome?

A standard enrichment method employs Fisher's exact test, a statistical test that evaluates a 2x2 contingency table (in this case, number of genes in my set *versus* number of genes from genome not in my set, and number of genes with GO term Z

versus number of genes without term Z). This test produces a p-value between 0 and 1, where  $p \leq 0.05$  is considered significant (that is, less than 5% probability that the enrichment is due to chance). However, the test is performed for each of the 100s of GO terms, increasing the chances that a GO term will be incorrectly considered enriched (a false positive, or type I, error). Thus, the original p-value must be adjusted for so-called multiple hypothesis testing, resulting in an adjusted p-value such as the Benjamini-Hochberg false discovery rate (FDR) or Bonferroni adjusted p-value.

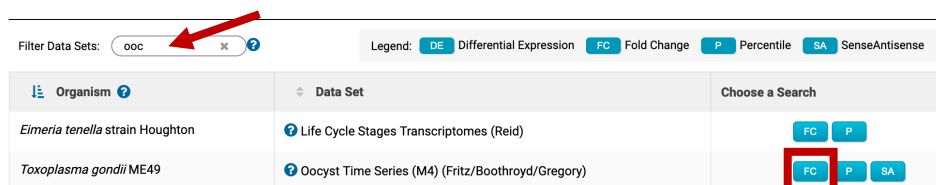
1. In order to run a GO enrichment analysis, you need a list of genes to test. This can be a list of gene IDs from your experimental results that you can upload using the ID search or a gene list resulting from a search you conducted on a VEuPathDB website. For this example, in ToxoDB, we will identify genes that are differentially regulated over time.

- a. Navigate to the RNA-Seq searches and find the data set called “**Oocyst Time Series (M4)**” from Fritz *et al.* A quick way of getting to the RNA-Seq searches is to type ‘rna’ in the filter box on the left of the home page and click on the RNA-Seq Evidence link. See image below.



- b. The RNA-Seq evidence page includes a list of all data sets that are loaded in the website. To quickly find a dataset, you can start typing key words in the “Filter Data Sets” box. For example, start typing the word “oocyst”.

### Identify Genes based on RNA-Seq Evidence



- c. Once you find the data set of interest, click on the fold-change (FC) option. This will open a search page that contains the parameters that you can manipulate to search this data set. For this exercise, identify genes that are upregulated by 20-fold in the day 4 and day 10 time points compared to the day 0 time point. Parameters to set:
1. Up-regulated
  2. 20-fold
  3. Maximum
  4. Day 0
  5. Minimum
  6. Day 4 and 10

#### Identify Genes based on T. gondii ME49 Oocyst Time Series (M4) RNA-Seq (fold change)

For the Experiment  
Oocyst Time Series (M4) - Sense

return protein coding genes  
that are up-regulated  
with a Fold change  $\geq$  20  
between each gene's maximum expression value  
(or a Floor of 10 reads)  
in the following Reference Samples

☒ day 0  
☐ day 4  
☐ day 10

select all | clear all

and its minimum expression value  
(or the Floor selected above)  
in the following Comparison Samples

☐ day 0  
☒ day 4  
☒ day 10

select all | clear all

**Example showing one gene that would meet search criteria**  
(Dots represent this gene's expression values for selected samples)

For each gene, the search calculates:

$$\text{fold change} = \frac{\text{minimum expression value in comparison}}{\text{reference expression value}}$$

and returns genes when fold change  $\geq$  20.

You are searching for genes that are **up-regulated** between one **reference sample** and at least two **comparison samples**.

This calculation creates the **narrowest** window of expression values in which to look for genes that meet your fold change cutoff. To broaden the window, use the average or maximum comparison value.

Get Answer

- d. Once you have set the parameters, click the “Get Answer” button at the bottom of the search. This will return a one-step search strategy. How many genes did you get?

2. To run a GO enrichment analysis on these results, do the following:

- a. Click on the Analyze Results tab just above the list of genes (arrow in image below).

## My Search Strategies

[Opened \(1\)](#) All (1) Public (17) Help

Unnamed Search Strategy \*

TgM4 Oocyst RNA-Seq (fc)  
1,029 Genes

+ Add a step

Step 1

1,029 Genes (970 ortholog groups)

[Revise this search](#)

Gene Results Genome View **Analyze Results**

Organism Filter  
select all | clear all | expand all | collapse all  
☐ Hide zero counts  
Search organisms...  
☐ Eimeriidae 0

1 2 3 ... 52

Rows per page: 20

[Download](#) [Add to Basket](#) [Add Columns](#)

- b. Click on the “Analyze Results” tab to reveal the different analyses that you can run on your results. Besides GO enrichment, what other analyses are available?

Gene Results Genome View **New Analysis**

Analyze your Gene results with a tool below.

Gene Ontology Enrichment

Metabolic Pathway Enrichment

kinase  
phosphatase  
exported  
membrane  
Word Enrichment

- c. Click on the GO enrichment option. This will reveal the parameters that you can modify. For the purpose of this exercise, keep all the defaults and click on “Submit”.
- d. What is the top enriched GO term from this analysis?
- e. What do each of the columns in the analysis table represent? (Hint: move your mouse over the question mark next to each column header to get more information.)

Genes in your result with this term ?	Percent of bkgd genes in your result ?
Number of genes with this term in your result 2.	

- f. Try rerunning the GO enrichment analysis, but this time select the Molecular Function ontology. What is the top enriched GO term?

Gene Results Genome View Gene Ontology Enrichment\* x Gene Ontology Enrichment\* x Analyze Results

[Rename This Analysis | Duplicate]

### Gene Ontology Enrichment

Find Gene Ontology terms that are enriched in your gene result. [Read More](#)

▼ Parameters

**Organism** ? Toxoplasma gondii ME49

**Ontology** ?  
☐ Cellular Component  
☒ Molecular Function  
☐ Biological Process

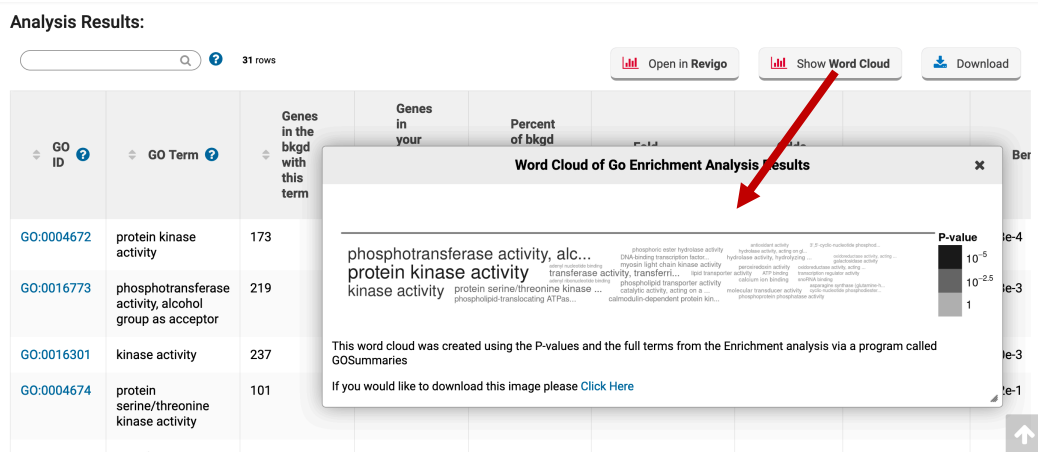
**Evidence** ?  
☒ Computed  
☒ Curated  
[select all](#) | [clear all](#)

**Limit to GO Slim terms** ? ☒ No  
☐ Yes

**P-Value cutoff** ? 0.05 (0 - 1)

Submit

- g. Click on the “Word Cloud” button above the analysis results. What does this do? (See image below).



**Additional resources:**

Gene Ontology:

<http://geneontology.org/docs/ontology-documentation/>

Enzyme Commission numbers:

<https://www.qmul.ac.uk/sbcs/iubmb/enzyme/>

More info on Fischer's exact test:

<http://www.biostathandbook.com/fishers.html>

Fisher's Exact Test and the Hypergeometric Distribution (the M&M example):

<https://youtu.be/udyAvvaMjfm>

Some more info about Odds ratios:

<http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2938757/>

False discovery rates and P value correction:

<http://brainder.org/2011/09/05/fdr-corrected-fdr-adjusted-p-values/>

GO Slim:

<http://www-legacy.geneontology.org/GO.slims.shtml>

REVIGO:

<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0021800>